

# Rat Anti-PEG IgM ELISA kit (MAK-0013)

This product is for research use only and is not intended for diagnostic use.

## PRODUCT INFORMATION

**Cat No.** MAK-0013

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**Storage** Indirect ELISA

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**Principle** This kit is for research use only. Under no circumstances should it be used for therapeutic or human diagnostic applications.

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**Intended Use** The attachment of polyethylene glycol chains to therapeutic biologic agents, a process referred to as PEGylation, prolongs the circulating half-life of the modified protein by slowing proteolytic degradation and by masking it from the immune system. However, repeated injections of PEGylated proteins can induce anti-PEG antibodies that increase the rate of clearance and thereby decrease efficacy (accelerated blood clearance, or ABC, phenomenon). To aid research, we have developed a rat anti-PEG IgM ELISA.

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**Introduction** The assay uses immobilized mono mPEGylated BSA (20 kDa PEG chain) as the capture antigen (coated on microtiter wells) and horseradish peroxidase (HRP) conjugated anti-rat IgM antibodies for detection. Serum or plasma samples are diluted and incubated alongside standards in the microtiter wells for 1 hour. The wells are subsequently washed, and HRP conjugate is added and incubated for 30 minutes. Anti-PEG IgM molecules are thus sandwiched between immobilized PEG and the detection antibody conjugate. The wells are then washed to remove unbound HRP-labeled antibodies, and TMB Reagent is added and incubated for 20 minutes at room temperature. This results in the development of a blue color. Color development is stopped by the addition of Stop Solution, changing the color to yellow, and optical density is measured spectrophotometrically at 450 nm. The concentration of anti-PEG IgM is proportional to the absorbance at 450 nm and is derived from a standard curve. This assay primarily detects antibodies directed against the polyoxyethylene

backbone of PEG. Studies at Mabioway Co., Ltd, in mice and rabbits have demonstrated that most anti-PEG antibodies induced by immunization with PEGylated proteins are directed against the backbone.

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**Principle of the assay** It has been reported that repeated injections of PEGylated proteins can induce anti-PEG antibodies that increase the rate of clearance and decrease drug efficacy (accelerated blood clearance, or ABC phenomenon). To aid research in this important area, we have developed a anti-PEG ELISA kit. Used for new drug research and development

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**Storage Note** The reference stock, HRP conjugate and the PEG-BSA coated plate should be stored at -20°C. All remaining kit components should be stored at 4°C. The microtiter plate should be kept in a sealed bag with desiccant. Kits will remain stable for six months from the date of purchase provided that the components are stored as described.

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**Materials** Materials provided with the kit:

- PEG-BSA coated plate (12 x 8-wells)
- Anti-Rat IgM HRP Stock (lyophilized)
- Reference Stock1 (lyophilized)
- 20x HRP PEG Wash: PEGW50-20, 50 ml
- HRP PEG Diluent: PEGD50-1, 50 ml
- TMB: TMB11-1, 11 ml
- Stop Solution: SS11-1, 11 ml

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**General instructions**

1. Please read and understand the instructions thoroughly before using the kit.
2. This kit is designed to measure anti-PEG IgM levels in serum collected approximately 7 days after immunization with PEG. Serum collected at post-immunization times greater than 7 days may contain elevated levels of anti-PEG IgG that compete with anti-PEG IgM for the immobilized PEG, thereby causing interference.
3. All reagents should be allowed to reach room temperature (25°C) before use.
4. The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.
5. Use only the wash solution and dilution buffer provided with the kit. PEG and PEGylated compounds are found in buffers conventionally used in ELISA's and cannot be used with this kit.
6. Kits are validated using plate shakers set at 150 rpm and 25°C. Performance of the assay at lower temperatures and/or mixing speeds will likely result in lower absorbance values.
7. Optimal results are achieved if, at each step, reagents are pipetted into the wells of the microtiter plate within 5 minutes.

<b>Diluent</b>	The diluent (PEGD50-1) is formulated for measurement PEG antibodies. It is supplied ready to use. DO NOT substitute other buffers.
<b>Sample Preparation</b>	<p>Studies at Mabioway Co., Ltd. indicate that anti-PEG IgM is undetectable in serum from control rats when tested at a 20-fold dilution. However, in serum from PEG-KLH immunized rats, levels of 845 to 47,940 u/ml (<math>16,488 \pm 15158</math>, mean <math>\pm</math> SD, n = 10) were observed seven days after immunization. Levels will vary with the immunization protocol and the PEG carrier protein. We suggest that samples initially be diluted 500-fold, but optimal dilutions must be determined empirically. A 500-fold dilution may be achieved as follows:</p> <ol style="list-style-type: none"> <li>1. Dispense 48 ul and 332.5 ul of diluent into separate tubes.</li> <li>2. Pipette and mix 2 ul of the serum/plasma sample into the tube containing 48 ul of diluent. This provides a 25-fold diluted sample.</li> <li>3. Mix 17.5 ul of the 25-fold diluted sample with the 332.5 ul of diluent in the second tube. This provides a 500-fold dilution.</li> </ol>
<b>HRP Anti-PEG Conjugate</b>	Approximately 15 minutes before needed, reconstitute the lyophilized HRP conjugate as described on the vial label and mix gently. Dilute as described on the vial label to give the working conjugate solution. The reconstituted conjugate stock must be stored at -20°C in a sealed vial if future use is intended.
<b>Wash Solution</b>	The wash solution is provided as a 20x stock. Prior to use, dilute the contents of the bottle (50 ml) with 950 ml of distilled or deionized water.
<b>Standards</b>	<ol style="list-style-type: none"> <li>1. The rat anti-PEG IgM standard is provided as a lyophilized. Reconstitute the stock as described on the vial label.</li> <li>2. Label 5 polypropylene or glass tubes as 100, 50, 25, 12.5, 6.25 and 3.125 u/ml.</li> <li>3. In the tube labeled 100 u/ml prepare the 100 u/ml standard as detailed on the stock vial label.</li> <li>4. Dispense 350 ul of diluent into the remaining tubes.</li> <li>5. Prepare a 50 u/ml standard by diluting and mixing 350 ul of the 100 u/ml standard with 350 ul of diluent in the tube labeled 50 u/ml.</li> <li>6. Similarly prepare the 25, 12.5, 6.25 and 3.125 u/ml standards by serial dilution.</li> </ol>
<b>Procedure</b>	<ol style="list-style-type: none"> <li>1. Secure the desired number of coated wells in the holder.</li> <li>2. Dispense 100 ul of standards and diluted samples into the wells (we</li> </ol>

- recommend that samples be tested in duplicate).
3. Incubate on an orbital micro-plate shaker at 150 rpm/25°C for 1 hour.
  4. Aspirate the contents of the microtiter wells and wash the wells 5 times with 1x wash solution using a plate washer (400 ul/well).
  5. Strike the wells sharply onto absorbent paper or paper towels to remove all residual wash solution.
  6. Add 100 ul of diluted HRP conjugate into each well.
  7. Incubate on an orbital micro-plate shaker at 150 rpm/25°C for 30 minutes.
  8. Wash as detailed above.
  9. Dispense 100 ul of TMB reagent into each well.
  10. Gently mix on an orbital micro-plate shaker at 150 rpm/25°C for 20 minutes.
  11. Stop the reaction by adding 100 ul of Stop Solution to each well.
  12. Gently mix. It is important to make sure that all the blue color changes to yellow.
  13. Read the optical density at 450 nm with a microtiter plate reader within five minutes<sup>2</sup>.

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## Important Tips

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## Results

1. Using curve fitting software, construct a standard curve by plotting absorbance values of the standards versus concentration.
2. Fit the standard curve to a second order polynomial model and determine the concentration of the samples from the standard curve.
3. Multiply the derived concentration by the dilution factor to determine the actual concentration in the samples.
4. If the A<sub>450</sub> values of samples fall outside the standard curve, samples should be diluted appropriately and re-tested.